

ScienceDirect



Dynamic and heterogeneous neural ensembles contribute to a memory engram

Brian M Sweis^{1,2}, William Mau¹, Sima Rabinowitz¹ and Denise J Cai¹



In the century since the notion of the 'engram' was first introduced to describe the physical manifestation of memory, new technologies for identifying cellular activity have enabled us to deepen our understanding of the possible physical substrate of memory. A number of studies have shown that memories are stored in a sparse population of neurons known as a neural ensemble or engram cells. While earlier investigations highlighted that the stability of neural ensembles underlies a memory representation, recent studies have found that neural ensembles are more dynamic and fluid than previously understood. Additionally, a number of studies have begun to dissect the cellular and molecular diversity of functionally distinct subpopulations of cells contained within an engram. We propose that ensemble fluidity and compositional heterogeneity support memory flexibility and functional diversity.

Addresses

 ¹ Icahn School of Medicine at Mount Sinai, Department of Neuroscience, New York, NY, 10029, United States
 ² Icahn School of Medicine at Mount Sinai, Department of Psychiatry,

New York, NY, 10029, United States

Corresponding author: Cai, Denise J (denise.cai@mssm.edu)

Current Opinion in Neurobiology 2021, 67:199-206

This review comes from a themed issue on Neurobiology of learning and plasticity

Edited by Sheena Josselyn and Tara Keck

https://doi.org/10.1016/j.conb.2020.11.017

0959-4388/© 2020 Elsevier Ltd. All rights reserved.

In the early twentieth century, Richard Semon introduced the term 'engram' to describe the physical manifestation of memory, defined as 'the enduring though primarily latent modification in the irritable substance produced by a stimulus' [1]. The biological basis for the engram was elusive, however, as early investigators were unable to find a specific engram within the cortex [2,3]. In recent years, new technologies for identifying and controlling cellular activity have enabled us to deepen our understanding of the possible physical 'trace' of memory. Several studies have shown that memories are stored in a sparse population of neurons, defined as engram cells or a neural ensemble [4–10]. While earlier studies highlighted that the stability of a neural ensemble underlies a stable memory representation, recent studies have suggested that neural ensembles are more fluid than previously thought [11–15]. Additionally, several groups have begun to dissect the cellular and molecular diversity of functionally distinct subpopulations of cells contained within an engram [16[•],17[•]]. We propose that memory ensembles comprise two additional properties that have only recently been explored: ensemble fluidity that supports memory flexibility and the compositional heterogeneity of subensembles that contributes differentially to memory functions.

Prior studies leveraged immediate-early gene tagging strategies as a way to identify which cells were activated during a learning or memory recall session [18]. By taking a 'snapshot' of which cells were activated during both learning and memory recall, investigators sought to find the engram-the physiological trace or storage site of the memory. In a study in the hippocampus using fluorescent *in-situ* hybridization measuring the expression of the immediate-early gene Arc, investigators found that many of the cells initially activated during encoding of an environment were reactivated when the animals reentered the environment 20 min later [19]. Similarly, another study found that amygdala cells initially activated during tone fear conditioning (tone paired with a shock) were likely to be reactivated when the animals heard the tone a week later, recalling the fear memory [20,21[•]]. Furthermore, the behavioral memory of the tone paired with the shock (assessed by the degree of freezing) was positively correlated with the amount of reactivation of the neural ensemble that was activated during initial learning. These studies suggest that memories are stored in neuronal ensembles and reactivation of those ensembles contributes to memory retrieval and subsequently to behavior.

If a memory is stored in a sparse neural ensemble, then silencing these cells should impair the brain's ability to retrieve the memory. Conversely, artificially activating these cells should induce the brain to retrieve the memory. To test the first hypothesis, an allocation strategy was used to bias a tone-fear memory to be stored in a subset of CREB+ cells in the amygdala. Later during recall when the tone was played, investigators silenced the CREB+ neurons (by ablating or temporarily silencing those neurons), which inhibited the ability of the animal to recall the tone-fear memory [21°,22°]. To test if artificially reactivating the neural ensemble was sufficient to recall a memory, an immediate-early gene tagging approach was used to tag hippocampal neurons that were activated during context conditioning (shock paired with a novel context) [23[•]]. Animals were later placed in a different and safe context, and when the neural ensemble tagged from context conditioning was artificially activated with an optogenetic strategy, the animals froze, suggesting that they were recalling at least some aspect of the fear memory. The memories stored in the ensembles were context specific, such that artificially activating a tagged ensemble of a neutral context (as a control) did not exhibit an expression of fear. Similar findings have been reported using a chemogenetic strategy [24]. Importantly, these findings have been reported for multiple types of memory [25–29] and across brain regions [9,30,31].

In addition to these studies, an extensive literature in hippocampal physiology also supports the theory that memories are stably encoded in neural ensembles. Using in vivo electrophysiology, early studies found that hippocampal neurons ('place cells') fire according to the animal's position in space [32–34]. Many studies have shown that place cells reliably fire when animals return to the same spatial location and this stability can be preserved across weeks, making these cells suitable for long-term storage of a cognitive map and possibly contributing important spatial information to specific engrams [34–37]. Critically, disruption of ensemble-specific reactivation of place cells (replay) impairs spatial memory [38]. Conversely, pairing intracranial stimulation with place cell reactivation during sleep can reinforce specific spatial memories and induce later awake spatially goaldirected behavior in mice [39]. Together, these studies strongly suggest that the memory engram exists in a stable neural ensemble and that this ensemble is necessary and sufficient for memory recall.

Memory engrams are flexibly updating with new information

While a number of *in vivo* electrophysiology studies have found hippocampal place cells to reliably fire again when animals return to the same spatial location, most of these studies compare place cell stability within one day because reliably recording from the same cells across multiple days has been challenging for electrophysiology. One study successfully recorded the same place cells across 2 days and found that place cells had more similar firing patterns within 6 hours than within 30 hours when the animal was exploring the same environment [40,41°]. Similar findings were reported using an *in vivo* imaging method which can record from the same cells across weeks [42°,43]. The probability that place cells would fire again when the animal was exposed to the same environment and was performing the same behavior decreased across weeks. Nevertheless, many of these place cells retained stable spatial information when they fired again days or weeks later. In other words, many place cells continued to fire with high fidelity in the same spatial location even though the probability of firing decreased across time. This ability to decode stable information in hippocampus despite changes in firing rates over time is not limited to spatial information but also extends to strategic information in rats trained on a rule-switching task [44[•]]. This phenomenon is known as representational 'drift,' where tuning is mostly stable while individual cell activity rates change across minutes to days. Representational drift has been reported across the brain [12].

Interestingly, the drift observed from in vivo data in place cell studies is consistent with immediate early gene results. While it is difficult to make direct comparisons across studies as the methods differ, it is worth noting that experiments using in vivo recording techniques found that when an animal returns to the same environment within minutes to hours the ensemble activity correlation of place cells is $\sim 90\%$ but drops to $\sim 60\%$ or lower across days $[40,42^{\circ},43]$. This decrease in reactivation of the same cells across time is generally consistent with observations from immediate early gene studies. When an animal returns to the same environment within 20 min, the ensemble reactivation in hippocampal CA1 is $\sim 90\%$ [19] but decreases to 30–50% days later [25,27]. While the exact reactivation rates differ across studies and methods, there seems to be consistency in that the relative reactivation rates of neural ensembles decrease across time.

This dynamic property of ensemble activity may provide a mechanism that allows certain features of a neuronal population to meaningfully track time while maintaining accurate tuning properties. One study supporting this theory showed time-dependent changes in spatial representations in the hippocampus [43]. Spatial tuning curves varied from day to day yet were sufficient to decode an animal's position along a linear track. Surprisingly, these activity patterns were also sufficient to decode the session in which the recording occurred. This finding led the authors to conclude that the variance in population activity could contain information about relative temporal distance between two similar encoding events [40,43].

Another possibility is that ensemble drift is not related to tracking time, but that instead, the function of dynamic ensembles might be to impose a strategy where neurons 'take turns' encoding new information to prevent too many memories from being allocated to the same population [Mau *et al.*, in press]. Long time intervals between encoding episodes, for example, can result in mostly non-overlapping neuronal ensembles $[45^\circ, 46^\circ]$. While the temporal distance between the two episodes could

theoretically be distinguished with this pattern of memory allocation, it is equally likely that temporal distance is not interpreted by a downstream reader these neurons project to as implied by Rubin et al. [43]. Instead, timedependent variance in ensembles could merely reflect the shift in priorities for which neurons receive incoming new memories, and the function of neuronal drift would be to distribute memories to neurons that were not recently recruited. This process could be beneficial for memory systems if plasticity is saturated in the population of neurons that make up an engram, which could occlude memory formation and memory-updating in these overworked populations [47]. To overcome this challenge, drift may support the turnover of ensembles to facilitate the availability of 'new cells' to encode new information to integrate with the prior memory or to encode a new, distinct memory altogether.

Endogenous changes and fluctuations in cellular activity may contribute to drift and serve as a mechanism for updating memories, linking those encoded close in time, while separating memories encoded at more distant time points [15,48], [Mau et al., in press]. One study focused on the hippocampus showed that 5 hours after context learning, the ensemble that encoded the memory had increased cellular excitability and this transient increase in intrinsic excitability allocated a second distinct context memory to be encoded by many of the same neurons as the first context memory [45°]. However, days later when excitability returned to basal levels, memory for a new context was no longer preferentially allocated to the neurons of the prior ensemble but was encoded instead in an independent ensemble of neurons. Sharing a neural ensemble between two memories functionally linked the two such that recall of one memory triggered recall of the temporally linked memory (encoded 5 hours apart, but not 24 hours apart, Figure 1). Similar findings have been observed in the lateral amygdala, where two different cued fear conditioning sessions administered 6 hours apart were more likely to be encoded by an overlapping population of neurons than those encoded a day apart [46[•]]. These memories were also shown to be behaviorally linked: extinguishing one fear memory also extinguished the other. While the studies described here focused on how distinct memories can be linked during encoding, other studies have demonstrated that transient increases in intrinsic excitability in ensemble cells can also occur during memory retrieval [49], priming the retrieved memory to be updated with new information [50].

Similar findings have been reported in human studies investigating how memory representations are linked across time. Functional imaging in humans revealed that neural representations for object pairs heavily overlapped if spaced 30 min apart, but not 24 hours apart [51]. Within the same temporal block of imaging, hippocampal activity patterns are more similar than when separated by an intervening event, suggesting that episodes are segregated based on time [52]. Furthermore, fear from aversive memories can transfer to neutral memories if encoded close in time [53,54]. Taken together, these findings across species, techniques, and behaviors suggest that endogenous changes and fluctuations in cellular excitability across time and experience can support the linking and updating of memories (Figure 1).

Memory engrams are heterogeneous in their composition

Earlier work on memory focused on engrams consisting mostly of excitatory neurons. Recent evidence, however, points to greater functional heterogeneity within ensembles than previously thought. Heterogeneous population activities have been observed during various types of learning. Diversity in neural firing dynamics in the hippocampus has been proposed to reflect the familiarity or novelty of learned information. One study found that familiarity was encoded by fast-firing, less-modifiable neurons where novel features of an experience were represented by a different set of slowly firing and highly plastic cells [55]. Another study investigated how hippocampal ensembles differentially represent context and space [56]. Similar to prior studies, Tanaka et al. found that a subset of CA1 cells expressed c-Fos after an animal experienced a novel context (A) and many of these cells were reactivated when the animal returned to the same context (A) but not when the animal explored another novel context (B), demonstrating context specificity of a neural ensemble. Interestingly, the cells that expressed c-Fos and reactivated during memory recall when revisiting context A did not fire in the same spatial location as during the initial memory encoding of this context. Surprisingly, between visits, the place fields of these cells had 'remapped' to a different spatial location within the same context. Non-c-Fos cells also, surprisingly, showed more stable spatial coding, as they were more likely to fire in the same spatial location during the recall session in context A. These data suggest that distinct hippocampal ensembles may store spatial and contextual information. Recent studies have also shown that subensembles contain different components of a memory, which are then orchestrated to constitute a memory [57-59]. In the hippocampus, hierarchical structures can organize relational information of multiple features of the environment [60].

While identifying subensembles is gaining increasing attention in the study of memory engrams, it is also important to highlight that distinct ensembles can encode the same information. For instance, multiple hippocampal ensembles can encode the same spatial context [61]. Elucidating both the divergent roles of subensembles, as well as the convergent roles of separate ensembles, will be critically important to understand the full complexity of





Neural ensembles are temporally dynamic.

A population of neurons whose firing patterns are tied to the encoding of and/or retrieval of a specific event in time is thought to comprise a memory trace, or engram. A sparse population of neurons that represent Event A are shaded in green and are distinct from the neurons that represent a separate episode occurring days later, Event B, shaded in red. Separate memory episodes that occur close in time to one another are more likely to share overlapping populations of neurons, as depicted in the cells shaded in blue corresponding to Event C occurring hours after Event B. This phenomenon is known as temporal memory-linking.

engrams. Very few studies have dissected how functionally distinct neuronal ensembles can be distinguished within an engram at the molecular and cellular levels. Genetically encoded activity reporters based on immediate early genes, such as c-Fos and Arc, commonly considered proxies for neuronal activity, have been used to identify neuronal ensembles in engrams. Until very recently, most studies have focused on ensembles defined by a single activity-dependent pathway [18]. However, activity-dependent pathways are known to be highly diverse: they respond differently to external stimuli and mediate distinct cellular and synaptic processes [62,63]. For instance, it is known that Arc proteins have virus-like capsid capabilities that can influence synaptic communication between neurons and underestimating this feature of Arc expressing neurons may miss critical network architecture features of how ensemble microcircuitry forms and is maintained [63,64]. Additionally, a recent study dissected two molecularly and functionally distinct ensembles within the dentate gyrus of the hippocampus underlying a contextual fear memory engram. Neurons that expressed c-Fos at learning, which mediates long-term potentiation of excitatory synapses, supported memory generalization, while neurons that expressed Npas4 at learning, which preferentially recruits inhibitory synapses onto excitatory neurons, regulated memory discrimination [17[•]] (Figure 2). The two molecularly distinct ensembles were both reactivated during recall, playing subtle but important roles in balancing memory generalization versus discrimination. These types of approaches to identifying how molecularly distinct cellular profiles can be linked to different physiological and behavioral functions have been appreciated in other areas of research. Bringing such molecular insights to the engram field, particularly as newer technologies combining

Current Opinion in Neurobiology 2021, 67:199-206

imaging and transcriptomics tools emerge, will advance systems neuroscience at a more interdisciplinary level [65].

Most studies on memory engrams have focused on the storage of memories in a neural ensemble consisting of excitatory neurons hypothesized to store stimulus associations through persistent changes in excitatory synapse

Figure 2



Neural ensembles consist of heterogenous subpopulations. Neurons belonging to an ensemble that represents a specific memory may be composed of distinct subpopulations of cells defined by molecular composition, circuit-specificity, and/or functional output. Different immediate-early genes (IEG, e.g. c-Fos, Npas4, Arc) that are commonly used to tag neurons tied to a specific event in an activitydependent manner can label separate subpopulations of neurons that, for example, receive distinct inputs and are important for fundamentally different behaviors, shaded in magenta and blue, respectively. strength and density [4,58,66]. In contrast, GABAergic interneurons are generally thought to inhibit excitatory neurons, and have been suggested to constrain ensemble size and to modulate memory strength and the specificity of learning [46°,67–70]. While several studies show that interneurons play an important supporting role in memory storage, recent evidence demonstrates that interneurons can also have a direct role in storing memories through their own functional plasticity. One study has shown that a subset of prefrontal somatostatin (SST) interneurons was activated during initial learning of tone-shock pairing and this ensemble of SST cells also exhibited enhanced plasticity (as shown by enhanced synaptic transmission) [16[•]]. Inactivating this specific SST ensemble reduced memory recall, while activating the SST ensemble elicited memory recall. This phenomenon was specific to SST neurons and was not seen in other interneurons (e.g. parvalbumin interneurons). Emerging evidence suggests that a memory engram contains different cell types and signaling pathways to engage different synaptic and circuit mechanisms to modulate memory-guided behaviors [71].

Conclusion and future directions

With the advent of more advanced experimental tools, the cellular and molecular properties of engrams can today be characterized in unprecedented ways [72-74]. We are discovering that engrams are more dynamic and fluid in their population codes than previously understood. Rather than mere noise in the system, these dvnamic properties may support complex computational capacities that might be useful for updating and integrating new information with existing memories across time. Furthermore, it is becoming increasingly clear that the cellular makeup of an engram's constituent neurons is more heterogenous than it appears, contributing to more complex microcircuitry that needs further characterization. Developing newer analysis methods of neural activity using unsupervised approaches can help us identify clusters of neural activity by how they are internally related in the brain rather than by experimenter-imposed behavioral labels [75]. Such knowledge, when combined with other approaches, may reveal unexpected functional roles of population activity as well as the how different ensembles or subensembles of cells may contribute to a function. In addition to analysis tools, developing more sensitive approaches with increased spatial and temporal resolution when recording, tagging and controlling neural activity, including newer advancements in optogenetics, chemogenetics, multi-channel calcium imaging, activity-dependent engineering strategies, and holography technologies will allow for a more refined understanding of the importance of spatiotemporal dynamics in memory processing [76-81]. Development of more sensitive behavioral readouts of memory representations will also help to further engram research. Much of the engram literature to date has concentrated on fear conditioning related behaviors, which, while useful for their simplicity, are limited by simple behavioral readouts. Increasing the complexity of the behavioral tasks, as well as the ability to extract more subtle behaviors from existing paradigms to probe the functional diversity of memory representations will be needed to advance the engram field [82]. Improving our knowledge of the complexity of the temporal and cellular properties of memory engrams will enable to us to better understand the multifaceted nature of memory representations and how they contribute to behavioral outcomes.

Conflict of interest statement

Nothing declared.

CRediT authorship contribution statement

Brian M Sweis: Conceptualization, Writing - original draft, Writing - review & editing. William Mau: Conceptualization, Writing - review & editing. Sima Rabinowitz: Conceptualization, Writing - review & editing. Denise J Cai: Conceptualization, Writing - original draft, Writing review & editing, Project administration, Funding acquisition.

Acknowledgements

We would like to thank Yosif Zaki, Natasha Berryman and Zhe Dong for insightful comments on a prior version of this manuscript. This work was funded by and N.I.H.F32 AG067640 to WM; NIH R01 MH120162, NIH DP2MH122399, One Mind Otsuka Rising Star Award, McKnight Memory and Cognitive Disorders Award, Klingenstein-Simons Fellowship Award in Neuroscience, Mount Sinai Distinguished Scholar Award, Brain Research Foundation Award and NARSAD Young Investigator Award to DJC.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- 1. Semon RW, Simon L: *The Mneme*. London, New York: G. Allen & Unwin Ltd.; The Macmillan Company; 1921.
- Lashley KS: Integrative functions of the cerebral cortex. Physiol Rev 1933, 13:0001-0042.
- Lashley KS: Studies of Cerebral Function in Learning. XI. The Behavior of the Rat in Latch Box Situations. The Mechanism of Vision. XII. Nervous Structures Concerned in the Acquisition and Retention of Habits Based on Reactions to Light. Baltimore: The Johns Hopkins Press; 1935.
- 4. Josselyn SA, Tonegawa S: Memory engrams: recalling the past and imagining the future. *Science* 2020, 367.
- Josselyn SA, Kohler S, Frankland PW: Heroes of the engram. J Neurosci 2017, 37:4647-4657.
- 6. Josselyn SA, Kohler S, Frankland PW: Finding the engram. *Nat Rev Neurosci* 2015, **16**:521-534.
- Poo MM, Pignatelli M, Ryan TJ, Tonegawa S, Bonhoeffer T, Martin KC, Rudenko A, Tsai LH, Tsien RW, Fishell G *et al.*: What is memory? The present state of the engram. *BMC Biol* 2016, 14:40.
- Tonegawa S, Liu X, Ramirez S, Redondo R: Memory engram cells have come of age. Neuron 2015, 87:918-931.

- Rogerson T, Jayaprakash B, Cai DJ, Sano Y, Lee YS, Zhou Y, Bekal P, Deisseroth K, Silva AJ: Molecular and cellular mechanisms for trapping and activating emotional memories. *PLoS One* 2016, 11:e0161655.
- Frankland PW, Josselyn SA, Kohler S: The neurobiological foundation of memory retrieval. Nat Neurosci 2019, 22:1576-1585.
- 11. Rule ME, Loback AR, Raman DV, Driscoll LN, Harvey CD, O'Leary T: **Stable task information from an unstable neural population**. *eLife* 2020, **9**.
- Rule ME, O'Leary T, Harvey CD: Causes and consequences of representational drift. Curr Opin Neurobiol 2019, 58:141-147.
- Driscoll LN, Pettit NL, Minderer M, Chettih SN, Harvey CD: Dynamic reorganization of neuronal activity patterns in parietal cortex. *Cell* 2017, 170:986-999 e916.
- Sehgal M, Zhou M, Lavi A, Huang S, Zhou Y, Silva AJ: Memory allocation mechanisms underlie memory linking across time. Neurobiol Learn Mem 2018, 153:21-25.
- Chen L, Cummings KA, Mau W, Zaki Y, Dong Z, Rabinowitz S, Clem RL, Shuman T, Cai DJ: The role of intrinsic excitability in the evolution of memory: significance in memory allocation, consolidation, and updating. *Neurobiol Learn Mem* 2020, 173:107266.
- Cummings KA, Clem RL: Prefrontal somatostatin interneurons
 encode fear memory. Nat Neurosci 2020, 23:61-74

This study was one of the first to show a direct role of a GABAergic interneuron ensemble to store a specific memory, and this ensemble was necessary and sufficient for encoding and retrieving a fearful memory. These authors found that synaptic potentiation of cue-responsive somatostatin interneurons in the prefrontal cortext can disinhibit cortical projection neurons to recruit remote brain regions involved in fear responses.

17. Sun X, Bernstein MJ, Meng M, Rao S, Sorensen AT, Yao L,

 Zhang X, Anikeeva PO, Lin Y: Functionally distinct neuronal ensembles within the memory engram. *Cell* 2020, 181:410-423 e417

This study was one of the first to show that a memory engram consists of molecularly distinct subensembles that can differentially contribute to memory function. These authors found that dentate gyrus engrams defined by Fos-dependent versus Npas4-dependent transcriptional pathways undergo unique synaptic changes after contextual fear conditioning and promote memory generalization versus discrimination, respectively.

- Mayford M, Reijmers L: Exploring memory representations with activity-based genetics. Cold Spring Harb Perspect Biol 2015, 8: a021832.
- Guzowski JF, McNaughton BL, Barnes CA, Worley PF: Environment-specific expression of the immediate-early gene Arc in hippocampal neuronal ensembles. Nat Neurosci 1999, 2:1120-1124.
- Reijmers L, Mayford M: Genetic control of active neural circuits. Front Mol Neurosci 2009, 2:27.
- Han JH, Kushner SA, Yiu AP, Hsiang HL, Buch T, Waisman A, Bontempi B, Neve RL, Frankland PW, Josselyn SA: Selective

erasure of a fear memory. Science 2009, **323**:1492-1496 This study was one of the first to show that selectively killing the cells that encoded a fear memory persistently inhibited recall of the memory. Using an inducible diptheria-toxin strategy, these authors ablated a population of neurons in the lateral amygdala overexpressing CREB preferentially recruited into a memory trace.

Zhou Y, Won J, Karlsson MG, Zhou M, Rogerson T, Balaji J,
 Neve R, Poirazi P, Silva AJ: CREB regulates excitability and the allocation of memory to subsets of neurons in the amygdala. Nat Neurosci 2009, 12:1438-1443

This study was one of the first to show that temporarily silencing the cells that encoded a fear memory inhibited the recall of the memory. These authors found that lateral amygdala cells with higher CREB levels show increased excitability and synaptic changes after fear conditioning and that virally manipulating CREB can modulate the allocation of fear memories to specific cells.

- 23. Liu X, Ramirez S, Pang PT, Puryear CB, Govindarajan A
- Deisseroth K, Tonegawa S: Optogenetic stimulation of a

hippocampal engram activates fear memory recall. Nature 2012, 484:381-385

This was one of the first studies to show that optogenetically activating the hippocampal ensemble that encoded a fear memory was sufficient to induce recall of that fear memory in a context-specific manner.

- Garner AR, Rowland DC, Hwang SY, Baumgaertel K, Roth BL, Kentros C, Mayford M: Generation of a synthetic memory trace. Science 2012, 335:1513-1516.
- 25. Ramirez S, Liu X, MacDonald CJ, Moffa A, Zhou J, Redondo RL, Tonegawa S: Activating positive memory engrams suppresses depression-like behaviour. *Nature* 2015, **522**:335-339.
- Redondo RL, Kim J, Arons AL, Ramirez S, Liu X, Tonegawa S: Bidirectional switch of the valence associated with a hippocampal contextual memory engram. *Nature* 2014, 513:426-430.
- 27. Ramirez S, Liu X, Lin PA, Suh J, Pignatelli M, Redondo RL, Ryan TJ, Tonegawa S: Creating a false memory in the hippocampus. *Science* 2013, 341:387-391.
- Denny CA, Kheirbek MA, Alba EL, Tanaka KF, Brachman RA, Laughman KB, Tomm NK, Turi GF, Losonczy A, Hen R: Hippocampal memory traces are differentially modulated by experience, time, and adult neurogenesis. *Neuron* 2014, 83:189-201.
- Trouche S, Perestenko PV, van de Ven GM, Bratley CT, McNamara CG, Campo-Urriza N, Black SL, Reijmers LG, Dupret D: Recoding a cocaine-place memory engram to a neutral engram in the hippocampus. Nat Neurosci 2016, 19:564-567.
- Cowansage KK, Shuman T, Dillingham BC, Chang A, Golshani P, Mayford M: Direct reactivation of a coherent neocortical memory of context. Neuron 2014, 84:432-441.
- Vetere G, Tran LM, Moberg S, Steadman PE, Restivo L, Morrison FG, Ressler KJ, Josselyn SA, Frankland PW: Memory formation in the absence of experience. *Nat Neurosci* 2019, 22:933-940.
- O'Keefe J, Dostrovsky J: The hippocampus as a spatial map. Preliminary evidence from unit activity in the freely-moving rat. Brain Res 1971, 34:171-175.
- **33.** Wilson MA, McNaughton BL: **Dynamics of the hippocampal ensemble code for space**. *Science* 1993, **261**:1055-1058.
- 34. Eichenbaum H: On the integration of space, time, and memory. Neuron 2017, 95:1007-1018.
- Schiller D, Eichenbaum H, Buffalo EA, Davachi L, Foster DJ, Leutgeb S, Ranganath C: Memory and space: towards an understanding of the cognitive map. *J Neurosci* 2015, 35:13904-13911.
- Foster DJ: Replay comes of age. Annu Rev Neurosci 2017, 40:581-602.
- Thompson LT, Best PJ: Long-term stability of the place-field activity of single units recorded from the dorsal hippocampus of freely behaving rats. *Brain Res* 1990, 509:299-308.
- Gridchyn I, Schoenenberger P, O'Neill J, Csicsvari J: Assemblyspecific disruption of hippocampal replay leads to selective memory deficit. *Neuron* 2020, 106:291-300 e296.
- de Lavilleon G, Lacroix MM, Rondi-Reig L, Benchenane K: Explicit memory creation during sleep demonstrates a causal role of place cells in navigation. Nat Neurosci 2015, 18:493-495.
- Mankin EA, Sparks FT, Slayyeh B, Sutherland RJ, Leutgeb S, Leutgeb JK: Neuronal code for extended time in the hippocampus. Proc Natl Acad Sci U S A 2012, 109:19462-19467.
- 41. Mankin EA, Diehl GW, Sparks FT, Leutgeb S, Leutgeb JK:
- Hippocampal CA2 activity patterns change over time to a larger extent than between spatial contexts. *Neuron* 2015, 85:190-201

This study was one of the first to use *in vivo* physiology to demonstrate 'driff' in place cells across days. These authors found that activity of hippocampal CA1 cells, but not CA3 cells, grow more different as time elapses despite maintaining contextual and spatial tuning.

42. Ziv Y, Burns LD, Cocker ED, Hamel EO, Ghosh KK, Kitch LJ, El Gamal A, Schnitzer MJ: Long-term dynamics of CA1 hippocampal place codes. Nat Neurosci 2013, 16:264-266

This study was one of the first to use in vivo calcium imaging to demonstrate temporal 'drift' in place cells across weeks. These authors tracked thousands of cells and found that the odds hippocampal CA1 pyramidal neurons recur with the same place field across any two days is ~25%, revealing much more dynamic remapping than previously thought.

- Rubin A, Geva N, Sheintuch L, Ziv Y: Hippocampal ensemble dynamics timestamp events in long-term memory. *eLife* 2015, 4.
- Hasz BM, Redish AD: Dorsomedial prefrontal cortex and
 hippocampus represent strategic context even while simultaneously changing representation throughout a task session. Neurobiol Learn Mem 2020, 171:107215

This study found that both hippocampal and prefrontal ensembles demonstrate 'drift' over time not only in representations of place but also in representation of strategy on a rule-switching task. These authors found that both regions could maintain task contingency representations despite 'drift', but that prefrontal ensembles 'drift' faster than in hippocampus.

- 45. Cai DJ, Aharoni D, Shuman T, Shobe J, Biane J, Song W, Wei B,
 Veshkini M, La-Vu M, Lou J *et al.*: A shared neural ensemble links
- Vesnkini M, La-Vu M, Lou J et al.: A shared neural ensemble links distinct contextual memories encoded close in time. Nature 2016, 534:115-118

This study was one of the first to show that a shared hippocampal ensemble linked contextual memories, such that fear from one contextual memory transferred to another neutral contextual memory encoded close in time. Using calcium imaging, these authors found that CA1 ensembles are more likely to overlap and link memories within a day but not across a week, a feature related to cellular excitability that is impaired in aged animals and could be rescued with excitatory DREADDs.

46. Rashid AJ, Yan C, Mercaldo V, Hsiang HL, Park S, Cole CJ, De
Cristofaro A, Yu J, Ramakrishnan C, Lee SY *et al.*: Competition between engrams influences fear memory formation and recall. *Science* 2016, 353:383-387

This study was one of the first to show that a shared amygdala ensemble linked memories, such that extinction of one fear memory led to extinction of another different fear memory. These authors found that optogenetically manipulating the excitability of lateral amygdala neurons could both coallocate overlapping populations for events close in time and disallocate nonverlapping populations for events separated in time.

- Choi JH, Sim SE, Kim JI, Choi DI, Oh J, Ye S, Lee J, Kim T, Ko HG, Lim CS et al.: Interregional synaptic maps among engram cells underlie memory formation. Science 2018, 360:430-435.
- Rogerson T, Cai DJ, Frank A, Sano Y, Shobe J, Lopez-Aranda MF, Silva AJ: Synaptic tagging during memory allocation. Nat Rev Neurosci 2014, 15:157-169.
- Pignatelli M, Ryan TJ, Roy DS, Lovett C, Smith LM, Muralidhar S, Tonegawa S: Engram cell excitability state determines the efficacy of memory retrieval. *Neuron* 2019, 101:274-284 e275.
- Yokose J, Okubo-Suzuki R, Nomoto M, Ohkawa N, Nishizono H, Suzuki A, Matsuo M, Tsujimura S, Takahashi Y, Nagase M et al.: Overlapping memory trace indispensable for linking, but not recalling, individual memories. Science 2017, 355:398-403.
- Zeithamova D, Preston AR: Temporal proximity promotes integration of overlapping events. J Cogn Neurosci 2017, 29:1311-1323.
- Ezzyat Y, Davachi L: Similarity breeds proximity: pattern similarity within and across contexts is related to later mnemonic judgments of temporal proximity. *Neuron* 2014, 81:1179-1189.
- Dunsmoor JE, Murty VP, Davachi L, Phelps EA: Emotional learning selectively and retroactively strengthens memories for related events. *Nature* 2015, 520:345.
- Yetton BD, Cai DJ, Spoormaker VI, Silva AJ, Mednick SC: Human memories can be linked by temporal proximity. Front Hum Neurosci 2019, 13:315.
- 55. Grosmark AD, Buzsaki G: Diversity in neural firing dynamics supports both rigid and learned hippocampal sequences. *Science* 2016, **351**:1440-1443.

- Tanaka KZ, He H, Tomar A, Niisato K, Huang AJY, McHugh TJ: The hippocampal engram maps experience but not place. Science 2018, 361:392-397.
- Ghandour K, Ohkawa N, Fung CCA, Asai H, Saitoh Y, Takekawa T, Okubo-Suzuki R, Soya S, Nishizono H, Matsuo M et al.: Orchestrated ensemble activities constitute a hippocampal memory engram. Nat Commun 2019, 10:2637.
- Goode TD, Tanaka KZ, Sahay A, McHugh TJ: An integrated index: engrams, place cells, and hippocampal memory. *Neuron* 2020, 107:805-820.
- 59. Tanaka KZ, McHugh TJ: The hippocampal engram as a memory index. J Exp Neurosci 2018, 12 1179069518815942.
- McKenzie S, Frank AJ, Kinsky NR, Porter B, Riviere PD, Eichenbaum H: Hippocampal representation of related and opposing memories develop within distinct, hierarchically organized neural schemas. *Neuron* 2014, 83:202-215.
- Sheintuch L, Geva N, Baumer H, Rechavi Y, Rubin A, Ziv Y: Multiple maps of the same spatial context can stably coexist in the mouse hippocampus. *Curr Biol* 2020, 30:1467-1476 e1466.
- Yap EL, Greenberg ME: Activity-regulated transcription: bridging the gap between neural activity and behavior. Neuron 2018, 100:330-348.
- 63. Pastuzyn ED, Day CE, Kearns RB, Kyrke-Smith M, Taibi AV, McCormick J, Yoder N, Belnap DM, Erlendsson S, Morado DR et al.: The neuronal gene Arc encodes a repurposed retrotransposon gag protein that mediates intercellular RNA transfer. Cell 2018, 173:275.
- Erlendsson S, Morado DR, Cullen HB, Feschotte C, Shepherd JD, Briggs JAG: Structures of virus-like capsids formed by the Drosophila neuronal Arc proteins. Nat Neurosci 2020, 23:172-175.
- Xu S, Yang H, Menon V, Lemire AL, Wang L, Henry FE, Turaga SC, Sternson SM: Behavioral state coding by molecularly defined paraventricular hypothalamic cell type ensembles. *Science* 2020, 370.
- Holtmaat A, Caroni P: Functional and structural underpinnings of neuronal assembly formation in learning. *Nat Neurosci* 2016, 19:1553-1562.
- Courtin J, Dejean C, Herry C: Prefrontal parvalbuminexpressing interneurons control fear behavior. Med Sci (Paris) 2014, 30:943-945.
- Siwani S, Franca ASC, Mikulovic S, Reis A, Hilscher MM, Edwards SJ, Leao RN, Tort ABL, Kullander K: OLMalpha2 cells bidirectionally modulate learning. *Neuron* 2018, 99:404-412 e403.
- Stefanelli T, Bertollini C, Luscher C, Muller D, Mendez P: Hippocampal somatostatin interneurons control the size of neuronal memory ensembles. *Neuron* 2016, 89:1074-1085.
- Grienberger C, Milstein AD, Bittner KC, Romani S, Magee JC: Inhibitory suppression of heterogeneously tuned excitation enhances spatial coding in CA1 place cells. Nat Neurosci 2017, 20:417-426.
- Barron HC, Vogels TP, Behrens TE, Ramaswami M: Inhibitory engrams in perception and memory. Proc Natl Acad Sci U S A 2017, 114:6666-6674.
- 72. Fenno LE, Ramakrishnan C, Kim YS, Evans KE, Lo M, Vesuna S, Inoue M, Cheung KYM, Yuen E, Pichamoorthy N et al.: Comprehensive dual- and triple-feature intersectional singlevector delivery of diverse functional payloads to cells of behaving mammals. Neuron 2020, 107:836-853 e811.
- Piatkevich KD, Bensussen S, Tseng HA, Shroff SN, Lopez-Huerta VG, Park D, Jung EE, Shemesh OA, Straub C, Gritton HJ et al.: Population imaging of neural activity in awake behaving mice. Nature 2019, 574:413-417.
- 74. Aharoni D, Khakh BS, Silva AJ, Golshani P: All the light that we can see: a new era in miniaturized microscopy. *Nat Methods* 2019, **16**:11-13.

- 75. Rubin A, Sheintuch L, Brande-Eilat N, Pinchasof O, Rechavi Y, Geva N, Ziv Y: Revealing neural correlates of behavior without behavioral measurements. *Nat Commun* 2019, **10**:4745.
- 76. Carrillo-Reid L, Yuste R: Playing the piano with the cortex: role of neuronal ensembles and pattern completion in perception and behavior. *Curr Opin Neurobiol* 2020, 64:89-95.
- Carrillo-Reid L, Han S, Yang W, Akrouh A, Yuste R: Controlling visually guided behavior by holographic recalling of cortical ensembles. *Cell* 2019, **178**:447-457 e445.
- Marshel JH, Kim YS, Machado TA, Quirin S, Benson B, Kadmon J, Raja C, Chibukhchyan A, Ramakrishnan C, Inoue M et al.: Cortical layer-specific critical dynamics triggering perception. Science 2019. 365.
- 79. Pegard NC, Mardinly AR, Oldenburg IA, Sridharan S, Waller L, Adesnik H: Three-dimensional scanless holographic

optogenetics with temporal focusing (3D-SHOT). Nat Commun 2017, 8:1228.

- Packer AM, Russell LE, Dalgleish HW, Hausser M: Simultaneous all-optical manipulation and recording of neural circuit activity with cellular resolution in vivo. Nat Methods 2015, 12:140-146.
- Robinson NTM, Descamps LAL, Russell LE, Buchholz MO, Bicknell BA, Antonov GK, Lau JYN, Nutbrown R, Schmidt-Hieber C, Hausser M: Targeted activation of hippocampal place cells drives memory-guided spatial behavior. *Cell* 2020, 183:1586-1599.e10.
- Mathis A, Mamidanna P, Cury KM, Abe T, Murthy VN, Mathis MW, Bethge M: DeepLabCut: markerless pose estimation of userdefined body parts with deep learning. *Nat Neurosci* 2018, 21:1281-1289.